Human milk oligosaccharides (HMOs) are a family of structurally diverse, complex sugars abundant in breast milk. HMO concentrations in human milk often exceed the concentration of all human milk proteins combined (Table 1). HMO profiles can vary based on genetics and geographic location, and can change over the course of lactation. HMOs are resistant to low pH and intestinal enzymes and are found in the urine and feces of breastfed infants. The oligosaccharides currently added to infant formula are structurally distinct from HMOs; however, some brands are beginning to include certain HMOs in newer formulas.

Table 1. Macronutrient composition in milk.

<table>
<thead>
<tr>
<th>Macronutrient, g/L</th>
<th>Human milk</th>
<th>Bovine milk</th>
</tr>
</thead>
<tbody>
<tr>
<td>Protein</td>
<td>12</td>
<td>35</td>
</tr>
<tr>
<td>Fat</td>
<td>35</td>
<td>35</td>
</tr>
<tr>
<td>Lactose</td>
<td>65</td>
<td>45</td>
</tr>
<tr>
<td>Oligosaccharides</td>
<td>5-15</td>
<td>0.05</td>
</tr>
</tbody>
</table>

HMOs act as prebiotics by providing a metabolic substrate for the growth of potentially beneficial bacteria. Bacterial communities in infant feces are able to grow when HMOs are provided as the only source of carbohydrates. Differences in microbial growth were seen with pooled HMO exposure versus individual HMOs; different responses were also seen with structurally different HMOs. The structure-dependent effects of HMOs depend on the infant and mother (e.g., “secretor status” of the mother can alter HMO composition in breast milk and HMO exposure in the infant).

In addition to providing “food for bugs,” accumulating evidence suggests HMOs have protective effects against infectious agents. HMOs have been shown to exhibit unique antimicrobial properties against group B Streptococcus, a leading cause of bacterial infection in newborns (Figure 1). The bacteriostatic activity was limited to specific non-sialylated HMOs (i.e., lacto-N-tetraose) and synergistic with other conventional antibiotic agents (i.e., vancomycin, ciprofloxacin). HMOs have also been shown to prevent infections by acting as antiadhesive antibacterials. HMOs isolated from pooled human milk significantly reduced attachment of enteropathogenic Escherichia coli (EPEC) to cultured epithelial cells and reduced EPEC colonization in suckling mice compared with untreated controls. A separate study demonstrated HMOs reduce attachment and cytotoxicity of the protozoan parasite Entamoeba histolytica when cultured with human intestinal epithelial cell lines.

HMOs may also have indirect effects on the infant microbiome by modulating epithelial and immune cell responses. Pre-treatment of bladder epithelial cells with HMOs reduced invasion of uropathogenic Escherichia coli (UPEC) and decreased host cell cytotoxicity. HMOs also reduced UPEC-mediated mitogen-activated

Figure 1. HMOs inhibit group B Streptococcus growth.3
protein kinase (MAPK) and nuclear factor-kappa B (NF-κB) activation, suggesting HMOs are protective against the proinflammatory effects of UPEC infection. Specific HMOs have been shown to directly affect immune responses by blocking macrophage inflammation; these HMOs are being tested in ongoing studies in animal models of chronic inflammation.

HMOs may play a role in preventing necrotizing enterocolitis (NEC), one of the most common and fatal gastrointestinal disorders in preterm infants. The risk of NEC is approximately 6- to 10-fold higher in formula-fed infants compared with breastfed infants; HMOs may contribute to the lower incidence of NEC in breastfed infants. To test this hypothesis, rat pups were orally gavaged with formula supplemented with or without pooled HMOs and exposed to hypoxia episodes. Formula supplemented with HMOs significantly improved 96-hour survival (95.0% vs 73.1%) and reduced NEC pathology scores (0.44 vs 1.98; \( P < 0.001 \)) in neonatal rats compared with formula alone. Further analyses demonstrated that a single HMO, disialyllacto-N-tetraose (DSLNT), had protective effects against NEC. A multicenter clinical cohort study was conducted to assess whether DSLNT can help prevent NEC in preterm infants. The HMO composition in breast milk from 200 mothers with very low birthweight infants was analyzed over the first 28 days postpartum. Among the 8 infants in the cohort who developed NEC (Bell stage 2 or 3), DSLNT concentrations were significantly lower in almost all milk samples compared with controls (ie, infants who did not develop NEC; Figure 2). Lower concentrations of DSLNT in breast milk could identify NEC cases prior to onset, suggesting DSLNT may be a potential noninvasive marker used to identify at risk infants and screen donor milk. Human intervention studies are needed to better define how DSLNT may prevent preterm infants from developing NEC. The same approach of combining preclinical studies and human cohort studies to inform clinical intervention studies is being applied to other diseases (eg, environmental enteric dysfunction and diarrheal disease, infant microbiome and obesity, allergies and asthma).

Figure 2. DSLNT concentrations are lower in breast milk from mothers with preterm infants who developed NEC.
HMO composition in breast milk is driven by several maternal factors, including genetics and possibly epigenetics, the environment (e.g., diet, lifestyle, smoking/drug exposure), health and disease, and parity. For example, high-fat diets are associated with less sialylated milk oligosaccharides in humans and mice, and HMO concentration has been shown to change with increase in parity. Additional studies are needed to determine how HMOs and lactation influence maternal health.

References


